Piperlongumine attenuates IL-1β-induced inflammatory response in chondrocytes.

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Abstract

Piperlongumine (PLM) is a natural product from the pods of a plant call *Piper longum* and was reported to possess antibacterial, antiangiogenic, neuroprotective, anti-inflammatory and anti-tumor activities. However, the role of PLM in inflammatory responses in human Osteoarthritis (OA) chondrocytes has not been yet explored. Thus, in this study, we investigated the anti-inflammatory action of PLM in human OA chondrocytes. Our results demonstrated that PLM treatment effectively reversed IL-1β-inhibited cell viability in a dose-dependent manner. In addition, PLM significantly inhibited the production of NO and PGE2, iNOS and COX-2 expression, as well as suppressed the production of MMP-3 and MMP-13 in IL-1β-stimulated human OA chondrocytes. Furthermore, PLM significantly prevented IL-1β-induced NF-κB activation in human OA chondrocytes. In conclusion, these findings demonstrated that PLM attenuated inflammatory responses in human OA chondrocytes stimulated by IL-1β, possibly through the NF-κB signaling pathway. Thus, PLM may serve as a potential anti-inflammatory agent in the treatment of OA.

Keywords: Osteoarthritis (OA), Piperlongumine (PLM), Chondrocytes, IL-1β.
Cell viability assay

Cell viability was assessed using the Cell Counting Kit-8 (CCK-8; Dojindo, Kumamoto, Japan) assay according to the manufacturer’s instructions. Briefly, after treatment, chondrocytes (1 × 10^4 cells/well) were plated into 96-well plates. Following culture for 24 h, 10 µl CCK-8 solution was added into each well and incubated at 37°C. After 3 h, the spectrophotometric absorbance was measured by an ELISA reader (Bio-Rad, Hercules, CA, USA) at 570 nm.

Detection of nitric Oxide (NO), prostaglandin E2 (PGE2) and MMP-3/13

NO production in the culture medium was detected by estimating the NO metabolite, nitrite, based on the Griess reaction as previously described [12]. The production of PGE2, MMP-3 and MMP-13 was measured by human ELISA PGE2, MMP-3 and MMP-13 kits (Sigma) according to the manufacturer’s respective instructions.

Quantitative real-time -PCR (qRT-PCR)

Total RNA was extracted from chondrocytes using TRIzol reagent (Invitrogen). Four microgram of total RNA for each sample was converted to cDNA by SuperScript reverse transcriptase and then amplified by Platinum Taq polymerase using the SuperScript One-Step RT-PCR kit (Invitrogen). The PCR primers for inducible Nitric Oxide Synthase (iNOS) were 5’-TTTCCAAGACACCTTCACCA-3’ (forward) and 5’-ATCTCCTTTGGTTACCGCTTCC-3’ (reverse); for cyclooxygenase-2 (COX-2) were 5’-GAGAGATGTATCCTCCCACAGTCA-3’ (forward) and 5’-GACCAGGCACCAGACCAAAG-3’ (reverse); and for GAPDH were 5’-ATGACAACTCCCTCAAGAT-3’ (forward) and 5’-GATCCACAACGGATACATT-3’ (reverse). Cycling conditions for amplification were: 95°C for 1 min; 40 cycles at 95°C for 45 s, 60°C for 30 s, and 72°C for 30 s; finally, 72°C for 5 min. The relative expression levels were calculated by 2^-ΔΔCt method and normalized to the expression of the housekeeping gene GAPDH.

Western blot

Human OA chondrocytes were homogenized and lysed with RIPA lysis buffer (Beyotime Biotechnology, Shanghai, China). Equal amounts of protein (30 µg) were loaded and separated by 12% SDS-PAGE, and transferred onto nitrocellulose membranes (Bio-Rad). After blocking with 5% non-fat milk in Tris-Buffered Saline (TBS) with 0.1% tween-20 (TBST) at room temperature for 1 h, the membranes were incubated with primary antibodies (anti-iNOS, anti-COX2, anti-phospho-NF-kB p65, anti-IκBα and anti-GAPDH) (Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) overnight at 4°C. Then, the membranes were incubated with Horseradish Peroxidase (HRP)-conjugated secondary antibodies (Santa Cruz Biotechnology) at room temperature for 1 h. Finally, protein bands on the membrane were visualized using enhanced chemiluminescence reagent (Invitrogen).

Statistical analysis

Experimental values are expressed as mean ± SD. Comparisons between two groups and among multiple groups were conducted using Student t-test and one-way analysis of variance followed by Tukey’s post-hoc test, respectively. Results were considered statistically significant at a P value less than 0.05.

Results

Effect of PLM on human OA chondrocyte viability

To examine the cytotoxicity of PLM on human OA chondrocytes, we performed a cell viability assay. As indicated in Figure 1A, PLM itself did not significantly affect the viability of chondrocytes up to 50 μg/ml. However, at a concentration of 100 μg/ml, a slight decrease in cell viability was observed. Next, we investigated whether PLM treatment could protect chondrocytes from IL-1β-induced cell death. As we expected, the viability of chondrocytes was significantly decreased by IL-1β. At the same time, we observed that pretreatment with PLM effectively reversed IL-1β-inhibited cell viability in a dose-dependent manner (Figure 1B).

Figure 1. Effect of PLM on human OA chondrocyte viability. A, Human chondrocytes (1 × 10^5 cells/well) were treated with various concentrations (0, 12.5, 25, 50 and 100 μg/ml) of PLM for 24 h, and cell viability was detected using the CCK-8 assay. B, Human chondrocytes (1 × 10^5 cells/well) were pretreated with or without various concentrations (12.5, 25 and 50 μg/ml) of PLM for 2 h and then incubated with 10 ng/ml IL-1β for 24 h, and cell viability was detected using the CCK-8 assay. Each value represents the mean ± SD of three independent experiments. *P<0.05 vs. control group, #P<0.05 vs. IL-1β group.
**PLM inhibits IL-1β-induced NO and PGE2 production in OA chondrocytes**

The next experiment was performed to examine the effects of PLM on NO and PGE2 production in IL-1β-stimulated human OA chondrocytes. As shown in Figure 2A, NO production was significantly increased in the IL-1β-treated group when compared with the control group. By contrast, PLM obviously inhibited NO production in a dose-dependent manner. Similarly, pretreatment with PLM also significantly suppressed PGE2 production induced by IL-1β in human OA chondrocytes (Figure 2B).

**Figure 2.** PLM inhibits IL-1β-induced NO and PGE2 production in OA chondrocytes. Human chondrocytes (1 × 10^5 cells/well) were pretreated with or without various concentrations (12.5, 25 and 50 μg/ml) of PLM for 2 h and then incubated with 10 ng/ml IL-1β for 24 h. A, NO production was detected by the Griess reaction. B, PGE2 production was measured by human ELISA PGE2 kit. Each value represents the mean ± SD of three independent experiments. *P<0.05 vs. control group, #P<0.05 vs. IL-1β group.

**PLM inhibits IL-1β-induced iNOS and COX-2 expression in OA chondrocytes**

Then, we investigated the effects of PLM on the expression of iNOS and COX-2 in chondrocytes exposed to IL-1β. The results of qRT-PCR analysis showed that compared to the control group, IL-1β treatment efficiently up-regulated the mRNA expression levels of iNOS and COX-2 in human OA chondrocytes; and these changes were suppressed by PLM (Figure 3A). In line with the results of qRT-PCR analysis, western blot analysis also demonstrated that the elevated protein expression levels of iNOS and COX-2 were reversed by PLM in IL-1β-stimulated chondrocytes (Figure 3B).

**Figure 3.** PLM inhibits IL-1β-induced iNOS and COX-2 expression in OA chondrocytes. Human chondrocytes (1 × 10^5 cells/well) were pretreated with or without various concentrations (12.5, 25 and 50 μg/ml) of PLM for 2 h and then incubated with 10 ng/ml IL-1β for 24 h. A, The mRNA expression levels of iNOS and COX-2 were evaluated using the qRT-PCR assay. B, The protein expression levels of iNOS and COX-2 were determined by western blot. Each value represents the mean ± SD of three independent experiments. *P<0.05 vs. control group, #P<0.05 vs. IL-1β group.

**PLM inhibits IL-1β-induced MMP-3 and MMP-13 production in OA chondrocytes**

MMPs play an important role in degrading cartilage. Therefore, herein, we investigated the effects of PLM on MMP-3 and MMP-13 production in chondrocytes stimulated by IL-1β. As shown in Figure 4, as compared with the control group, IL-1β stimulation significantly induced the production of MMP-3 and MMP-13, whereas pre-treatment with PLM significantly suppressed IL-1β-induced MMP-3 and MMP-13 production in OA chondrocytes (Figure 4B).
of MMP-3 and MMP-13 in human OA chondrocytes, which was efficiently prevented by PLM.

Figure 4. PLM inhibits IL-1β-induced MMP-3 and MMP-13 production in OA chondrocytes. Human chondrocytes (1 × 10^5 cells/well) were pretreated with or without various concentrations (12.5, 25 and 50 μg/ml) of PLM for 2 h and then incubated with 10 ng/ml IL-1β for 24 h. A, The production of MMP-3 was measured using the MMP-3 ELISA kit. B, The production of MMP-13 was evaluated by the MMP-13 ELISA kit. Each value represents the mean ± SD of three independent experiments. *P<0.05 vs. control group, #P<0.05 vs. IL-1β group.

PLM inhibits the activation of NF-κB signaling in OA chondrocytes

In order to explore whether the anti-inflammatory mechanism of PLM was associated with the inhibition of NF-κB activation, NF-κB signaling molecules were detected by western blot analysis. As indicated in Figure 5, IL-1β stimulation markedly increased the level of phospho-NF-κB p65, as well as induced IκBα degradation, as compared with the control group. However, PLM greatly inhibited IL-1β-induced NF-κB activation in human OA chondrocytes.

Figure 5. PLM inhibits the activation of NF-κB signaling in OA chondrocytes. Human chondrocytes (1 × 10^5 cells/well) were pretreated with or without various concentrations (12.5, 25 and 50 μg/ml) of PLM for 2 h and then incubated with 10 ng/ml IL-1β for 24 h. The levels of p-NF-κB p65 and IκBα were determined by western blot analysis. Protein levels were normalized to that of GAPDH. Each value represents the mean ± SD of three independent experiments. *P<0.05 vs. control group, #P<0.05 vs. IL-1β group.

Discussion

To our knowledge, this is the first report of the anti-inflammatory effects of PLM on human OA chondrocytes. Our results demonstrated that PLM treatment effectively reversed IL-1β-inhibited cell viability in a dose-dependent manner. In addition, PLM significantly inhibited the production of NO and PGE2, iNOS and COX-2 expression, as well as down-regulated the production of MMP-3 and MMP-13 in IL-1β-stimulated human OA chondrocytes. Mechanistic studies showed that PLM markedly suppressed IL-1β-induced NF-κB activation in human OA chondrocytes.

IL-1β has been widely used to mimic the microenvironment of OA for in vitro studies [13]. Thus, in this study, we used IL-1β-stimulated human OA chondrocytes to examine the anti-inflammatory effects of PLM on human OA chondrocytes. The results of this study showed that treatment of chondrocytes with IL-1β significantly decreased cell viability. This observation was consistent with the results of previous studies [14-16]. Additionally, pretreatment with PLM significantly reversed IL-1β-reduced cell viability. These findings suggest that the protective effect of PLM against pro-inflammatory responses induced by IL-1β in human OA chondrocytes.

Numerous studies have documented that overproduction of NO and PGE2 contribute to the development of OA [17-19]. NO is produced by iNOS in a variety of types of cells, including chondrocytes [20]. COX-2, known as the rating limiting enzyme, could produce the inflammatory mediator PGE2 in chondrocytes. In addition, IL-1β was found to up-regulate the expression of iNOS and COX-2, which lead to elevated production of NO and PGE2, respectively [21,22]. Besides, PLM has been reported to have anti-inflammatory properties. Lee et al. reported that PLM treatment dramatically inhibited
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Lipopolysaccharide (LPS)-induced the production of Tumor Necrosis Factor-α (TNF-α) and Interleukin (IL)-6 in Human Umbilical Vein Endothelial Cells (HUVECs) [23]. Consistent with the prior results, our findings showed that PLM inhibited the production of NO and PGE2, and the expression of iNOS and COX-2 in IL-1β-stimulated human OA chondrocytes. The above mentioned results suggest that PLM attenuated the IL-1β-induced chondrocyte injury through down-regulation of iNOS and COX-2 expression.

MMPs production and activation have been shown to play a critical role in the progression of OA [24,25]. MMP-13 is expressed mainly by articular chondrocytes, and preferentially cleaves the ECM components, such as type II collagen and aggrecan [26]. Besides, IL-1β is able to stimulate the synthesis of MMPs in chondrocytes [27]. In this study, we found that PLM obviously suppressed IL-1β-induced the production of MMP-3 and MMP-13 in chondrocytes. These data suggest that PLM may exhibit anti-OA effect through suppressing the production of MMPs.

Growing evidences showed that NF-κB is an important transcription factor and participates in the development of inflammatory diseases, including OA [28-30]. In response to IL-1β, NF-κB p65 dissociates from IκB and translocates into the nucleus to regulate the expression of inflammation-related genes, including MMP-1, 3, and 13, TNF-α, iNOS/NOS-2, PGE2 and COX-2 [31]. Previous studies indicated that stimulating of chondrocytes by IL-1β could induce NF-κB activation which subsequently regulating the expression of MMPs, COX-2, and iNOS [32,33]. In the present study, we showed that PLM markedly suppressed IL-1β-induced NF-κB activation in human OA chondrocytes. These results imply that the anti-inflammatory action of PLM on IL-1β-stimulated human OA chondrocytes was associated with the suppression of the NF-κB signaling pathway.

In conclusion, the results of this study demonstrated that PLM attenuated inflammatory responses in human OA chondrocytes stimulated by IL-1β, possibly through the NF-κB signaling pathway. Thus, PLM may serve as a potential anti-inflammatory agent in the treatment of OA.

References

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