In vitro study of novel degradable bone cement for osteoporotic compressive fractures.

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Abstract
A novel tricalcium phosphate Ca3(PO4)2 and calcium silicate CaSiO3 (TCP/CS) bone cement was developed and its mechanical properties and biocompatibility were evaluated from various proportions of TCP/CS composites which yield an injectable, self-solidified and degradable alternative biomaterial. The Osteoblasts (OB) were added into the TCP/CS composite disks and the properties of OB such as adhesion, proliferation, morphology and differentiation were observed. The solidification time of TCP/CS increases as the content of CS increases. The injectability of TCP/CS composite improves dramatically comparing that of TCP paste. The compressive strength of TCP/CS composites is greatly stronger than that of TCP alone. Furthermore, the degree of cell attachment, proliferation and differentiation, as well as biocompatibility significantly improves upon addition of CS into TCP. The TCP/CS composite with injectability possesses high strength and good biocompatibility, it might become a new biomaterial for vertebroplasty in the treatment of osteoporotic compressive fractures.

Keywords: Tricalcium phosphate, Calcium silicate, Biocompatibility, Biodegradability, Osteoporotic compressive fractures.

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Introduction
Osteoporotic Vertebral Compression Fractures (OVCFs) are the most common complications of osteoporosis [1-4]. Currently, minimally invasive vertebroplasty and balloon angioplasty are two main surgical treatments for OVCFs [5,6]. At present, Polymethylmethacrylate (PMMA) is the vertebral body forming material that have been most used in the world [7-10]. However, PMMA has many defects, for example, it cannot be absorbed in vivo, and present in the form of foreign matter in a long-term period to result in stress concentration between adjacent segments, and increase the risk of fractures of the adjacent spinal segments. It is simply a mechanical-supporting material without functions of bone conduction and induction, which hinders the healing of spinal fractures. Therefore, it is very important to develop an absorbable biomaterial that can accelerate bone forming for the clinical treatment of OVCFs.

Porous tricalcium phosphate TCP (Ca3(PO4)2) has been used in vertebroplasty, it possesses good biodegradability, biocompatibility and biological safety [1-8,11,12]. TCP has been investigated by scientists and engineers as a potential bone tissue engineering scaffold material [9-13]. However, it has not been widely applied clinically due to some shortcomings, such as rapid solidification, poor degradation and inconvenient operation [10,11,13]. CaSiO3 (CS) has a good ability to induce the deposition of hydroxyapatite, acceptable biocompatibility and bone binding and repair properties [14-17]. Recently, CS ceramics have been studied as bioceramics for new bone regeneration [18-20]. In this study, CS and TCP bone cement was prepared to improve the injectable properties and the biological activity of TCP cement and further provide a new strengthening biomaterial for OVCFs treatment.

Materials and Methods
Preparation of TCP/CS composites
Calcium hydroxide suspension was added drop wise into phosphoric acid solution under stirring at room temperature, the mixture was reacted for 4-5 h followed by filtering and drying at 80°C to prepare TCP powder. Porous TCP material was prepared from stearic acid particle filling. The spherical particles of stearic acid with high purity were divided into different mesh with sample sieve, the particles with appropriate particle size and TCP powder were mixed, Polyvinyl Alcohol (PVA) solution was added as a binder. After molding, stearic acid was molten at 80°C then dried to obtain the porous mould. The mould was placed in a sintering furnace, and heated to remove residual moisture, PVA, stearic acid and other residues. The prepared TCP ceramics were ground and mixed with CS powder according to different ratios. The powder mixtures
were calcined at 800°C for 2-3 h to produce TCP/CS nanocomposite powder. Polyethylene Oxide (PEO) was used as pore forming agent, TCP/CS powder and PEO particles were homogeneously mixed according to a mass ratio of 4:6, PVA solution was added as the binder to obtain biscuits. The biscuits were sintered at 800°C to give porous TCP/CS composites. The experimental grouping information is listed in Table 1.

Table 1. Experimental groups of TCP/CS composites.

<table>
<thead>
<tr>
<th>Group</th>
<th>TCP content (%)</th>
<th>CS content (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>90</td>
<td>10</td>
</tr>
<tr>
<td>B</td>
<td>80</td>
<td>20</td>
</tr>
<tr>
<td>C</td>
<td>70</td>
<td>30</td>
</tr>
<tr>
<td>TCP</td>
<td>100</td>
<td>0</td>
</tr>
</tbody>
</table>

Physical and chemical properties determination

The solidification time was determined by cement standard consistency setting time measuring instrument (Tianjin Qingda Instrument Equipment Co. Ltd.) according to a standard testing method. Six values were obtained with each group and the average value was calculated.

After mixing the solid and liquid phases of materials with different time, the mixtures were loaded into a disposable syringe needle and gradually extruded from the needle. The injectability was determined by the weight of replacing materials before and after extrusion. The fluidity (%) equals material weight before injection divided by weight of material before injection. Six values were obtained with each group and the average value was calculated.

A standard material sample is prepared by grinding. After curing for different time, the sample was loaded and pressed by a universal mechanical testing machine with a loading speed of 2 mm/min. The maximum fracture load ($F_{\text{max}}$) was recorded. The compressive strength is calculated by $F_{\text{max}}$ divided by the cross-sectional area of sample to evaluate its mechanical strength. Again, each group was measured 6 times, and the average value was calculated.

In vitro cytology experiment

The adhesion and proliferation of rat Osteoblasts (OB) on the surface of material were detected by MTT. The OBs were seeded on sterile materials from each group in a 24-well cell culture plate to culture for 5 and 10 d. The culture medium was transferred and washed 3 times. OBs were obtained by cleaving with 1% Triton-X100 solution and incubated for 24 h at 10°C. The activity of the enzyme in the pyrolysis product was determined by ALP kit, and the absorbance of the solution was measured at a wavelength of 405 nm. The total protein content of each cell was determined by BCA method. ALP activity was expressed using a standardized value for total protein.

Statistical method

Tongfang DA software (Version 2A.13B, Qinghua Tongfang Inc., China) was used for data processing. The experimental results were expressed by mean ± SD. Single-factor Analysis of Variance (ANOVA) was used to compare the difference between the control group and the experimental group, $p<0.05$ indicates that there are no significant differences between the data.

Results

Solidification time

The solidification times of materials in Groups A-C and control group are listed in Table 2. The solidification time of the sample material increases with the increase of CS content.

Table 2. The solidification times measured from experimental groups.

<table>
<thead>
<tr>
<th>Group</th>
<th>Solidification time (min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>54 ± 2.3</td>
</tr>
<tr>
<td>B</td>
<td>46 ± 1.8</td>
</tr>
<tr>
<td>C</td>
<td>35 ± 2.1</td>
</tr>
<tr>
<td>TCP</td>
<td>21 ± 0.8</td>
</tr>
</tbody>
</table>

Fluidity

The fluidity of materials in each material did not show significant difference upon mixing, they all possessed good injectability. After 10 min, the materials in Groups A-C can still maintain good fluidity, the injectability was obviously better than control group ($p<0.05$). After 20 min, the difference in injectability of each group further increased. The injectability of Group A which contains 30% CS reached 90%, comparing the value of 58% for the control group. After 30 min, the injection ability of all groups decreased to different extents, the control group decreased most, and the injectability was only 33% (Figure 1).
Figure 1. Fluidity of the materials in different experimental groups.

**Mechanical strength**

Table 3 lists the compressive strengths obtained from material samples in experimental groups. The compressive strength for each group increases with the increase of solidification time. At the same time point, the material containing 20% CS has the highest mechanical strength (76 MPa), significantly higher than the control group (32 MPa and 41 MPa at the 1\textsuperscript{st} and the 5\textsuperscript{th} d, respectively) (P<0.05).

**Table 3.** Comparison of compressive strengths obtained from different groups.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Compressive strength at the 1\textsuperscript{st} d (MPa)</th>
<th>Compressive strength at the 5\textsuperscript{th} d (MPa)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TCP/30% CS</td>
<td>64 ± 1.3</td>
<td>68 ± 1.6</td>
</tr>
<tr>
<td>TCP/20% CS</td>
<td>72 ± 1.9</td>
<td>76 ± 1.2</td>
</tr>
<tr>
<td>TCP/10% CS</td>
<td>51 ± 1.1</td>
<td>55 ± 0.9</td>
</tr>
<tr>
<td>TCP</td>
<td>32 ± 0.6</td>
<td>41 ± 0.5</td>
</tr>
</tbody>
</table>

**OBs adhesion ability determination on materials surface**

The results of MTT experiments showed that there was no significant difference between TCP control group and experimental groups after culturing for 5 h (P>0.05). After culturing for 10 h, the absorbance values (OD) of experimental groups were higher than TCP group with statistical difference (P<0.05), group C gave a highest value at 10 h (Figure 2).

**OB proliferation determination on materials surface**

As an increase in inoculation time, the cells exhibited a continuous increase on the surface of different materials. Figure 3 is proliferation curves of the cells. After culturing for 5 and 10 d, the OD value of all experimental groups were higher than the TCP group and the difference was statistically significant (P<0.05), suggesting that the materials in experimental groups are more conducive to the growth and proliferation of cells and they don’t have cell toxicity comparing the TCP group.

**Determination of osteogenic differentiation ability (ALP activity) of OBs**

As an increase of time, the ALP activity of cells in all groups gradually increased, the ALP activity detection data of cells and materials after culturing for 5 d and 10 d were listed in Table 4. The results showed that ALP activity of experimental groups was higher than that TCP group, the difference was statistically significant (P<0.05).

**Table 4.** Comparison of ALP activity in cells on materials surface.

<table>
<thead>
<tr>
<th>Groups</th>
<th>ALP activity at the 5\textsuperscript{th} d</th>
<th>ALP activity at the 10\textsuperscript{th} d</th>
</tr>
</thead>
<tbody>
<tr>
<td>TCP/30% CS</td>
<td>5.2</td>
<td>6.4</td>
</tr>
<tr>
<td>TCP/20% CS</td>
<td>4.8</td>
<td>6.2</td>
</tr>
<tr>
<td>TCP/10% CS</td>
<td>4.1</td>
<td>5.7</td>
</tr>
<tr>
<td>TCP</td>
<td>3.4</td>
<td>5.5</td>
</tr>
</tbody>
</table>
Discussion
At present, most of the vertebral filling materials for OVCFs treatment are Polymethylmethacrylate (PMMA). However, PMMA has many significant defects, such as simple mechanical support, no bone conduction and bone induction, cannot be absorbed and replaced by self-tissues; high elastic modulus, and even PMMA bone cement block loosening and displacement. Therefore, a new type of vertebroplasty replacement material is urgently needed to provide sufficient mechanical support for the osteoporotic vertebral body and promotes healing of bone fracture by bone conduction and bone induction.

TCP cement has been well developed thanks to its advantages, such as development, coagulation without releasing heat, and no cytotoxicity. TCP has been applied for vertebroplasty [6-12]. However, due to its brittleness, poor mechanical strength, curing difficulty after encountering body fluids and blood, this cement is difficult to meet the immediate stability requirements of elderly osteoporotic spine fracture after operation. Therefore it has been commonly used for young patients with type A vertebral fracture.

The use of PMMA bone cement filling to compression fracture of vertebral body is actually for the purpose of relieving pain from mechanical stability via bonding the tiny fracture block inside vertebral body [15,17,18]. However, this process also directly affects fracture healing. The results in this study showed that after co-culturing of cells with new alternative materials, alternative materials group was significantly higher than that of TPC group, suggesting that the introduction of CS materials helps to improve cell adhesion effect. The possible reason is that the SiO2 layer can be formed on the surface of the materials when it contacts with body fluid. It can produce high negative charge and promote cell adhesion to the material by adsorbing serum proteins. The SEM pictures showed that OBs can maintain normal cell morphology in the surface of TCP and the materials and adhere and grow through the protrusions in the material surface, suggesting that TCP and substitute materials are both suitable for cell growth, the good biological activity of new material can be confirmed.

After co-culturing the cells and testing materials for 5 and 10 d, ALP activity of experimental groups were higher than TPC group. ALP activity increased gradually with time, suggesting that addition of CS into TPC can improve the cell differentiation and biological activity better than TPC. In this study, the ions dissolved and precipitated on the surface of materials can also provide a Ca2+ and Si2+ rich microenvironment for cells, which is beneficial to the growth, proliferation and differentiation of cells. However, it has been reported that ALP activity in human bone marrow stromal cells is not significantly increased through bioglass [19]. But the in-vivo biological glass is able to greatly promote bone regeneration, the possible reason could be contributed to the complicated microenvironment in human body containing various bone cells and growth factors. Therefore, in-vivo bone regeneration promoting mechanism is more complicated, which is not only achieved by promoting cell differentiation of bone marrow stromal stem. Recent study also showed that a replacement material can effectively promote the differentiation of stromal cells of human bone marrow [20]. Therefore, there are still some differences about the promoting effect of CS on human bone marrow stromal stem cells. Future studies are needed to investigate the biological responses of alternative materials to stromal cells of human bone marrow.

Conclusions
A novel TCP/CS composite biomaterial with injectability was synthesized and developed in current study. This material possesses high strength and good biocompatibility, it might become a new biomaterial for vertebroplasty in the treatment of osteoporotic compressive fractures.

References


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