Evolution of cerebrovascular changes in mouse cortex during normal pregnancy by *in vivo* two-photon imaging.

Changbo Jin1,2,3#, Xinjia Han1,2,3#, Jinying Yang2, Huishu Liu1,2*
1Guangzhou Medical University, Guangzhou, PR China
2Department of Obstetrics, Guangzhou Women and Children's Medical Center, PR China
3Guangzhou Institute of Pediatrics, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou, PR China
#These authors contributed equally to this paper

**Abstract**

Past studies have shown that cerebral autoregulation is a physiologic process that maintains blood flow at an appropriate level. Impairment of cerebral autoregulation during pregnancy may contribute to further diseases like preeclampsia and eclampsia. However, little is known about the normative cerebrovascular adaptation during pregnancy. Two photon microscopy has become an important tool for living brain function research due to its deeper detection, higher resolution, lower photodamage compared to wide-field and confocal microscopy. In the present study we observed adaptive changes in the blood flow and vessel diameter from prepregnancy to late pregnancy by *in vivo* two-photon imaging. 12 female C57 mice were randomly divided into four groups: non-pregnancy, early-pregnancy, middle-pregnancy, late pregnancy. A thinned skull window was made over the motor cortex. One day later, Rhodamin B was injected intraperitoneally to label the vessels 15 minutes before beginning the imaging experiments. The animal was anesthetized and placed under two photon microscope to observe; blood flow signal pictures were taken using line-scan and analysed by blood flow analysis software programmed. The blood flow velocity of arteriovenous vessels and capillaries increased significantly in middle and late pregnancy compared to non-pregnancy and early-pregnancy; vascular diameters in middle and late pregnancy also became bigger than that in non-pregnancy and early-pregnancy. Both the blood flow velocity and diameter of cortical micro vessels increased significantly from non-pregnancy and early-pregnancy to middle and late pregnancy. Such results demonstrated a progressive cerebrovascular adaptation from prepregnancy to late pregnancy.

**Keywords:** Pregnancy, Blood flow velocity, Vascular diameters, Two photon.

**Accepted on April 24, 2017**

**Introduction**

Normal pregnancy is a state of physiological adaptation [1]. Changes in cardiovascular, renal, immune, and endocrine systems during pregnancy are necessary and important for maternal health, normal growth and development of the fetal-placenta unit [2]. Systemically, pregnancy is a high volume, low resistance state characterized by a large increase in cardiac output response to high levels of circulating hormones during the course of gestation [3]; Plasma volume also increases 40%-50% during pregnancy [4]. In addition, organs like uterus, kidney and heart undergo substantial increases in blood flow during pregnancy [5]. However, the adaptation of the brain to pregnancy is distinct from other organs. The brain is an organ of high metabolic demand that consumes 20% of the body's oxygen at rest, despite comprising only 2% of body weight [6]. Importantly, the brain has a relatively narrow capacity to tolerate changes in ion and water balance, and blood flow [6]. So the adaptation of the cerebral circulation during pregnancy appears to be to maintain a constant blood supply and the relative intolerance to increased blood volume. But the cerebral circulation is not unchanged during pregnancy.

Presently, the adaptations of the cerebral circulation to pregnancy and the related mechanism have not been understood clearly. It has been difficult to assess how CBF change during pregnancy in human. In recent studies, several non-invasive techniques such as Transcranial Doppler (TCD) ultrasound, Computed Tomography Perfusion (CTP), Magnetic Resonance Imaging (MRI), have been widely employed in patients to measure cerebral blood flow [7]. However, changes in blood flow calculated from TCD studies may not accurately reflect CBF due to the lack of information about vessel diameter [8]. In addition, all the above techniques have been
rarely used to study cerebral hemodynamics in experimental animals. Due to the nonlinear optical effects of two-photon absorption in biological tissue, the two photon microscopy technique allowed researchers to observe single vessels in the cerebral cortex of living brain [9]. Two photon imaging has a deeper penetration over several hundred micrometers of depth, higher resolution, more concentrated space focus, less tissue damage compared to laser confocal microscope and other conventional single photon microscopes.

During pregnancy, approximately 2-8% of women have some form of hypertensive disorder such as preeclampsia [10]. Preeclampsia accounts for 16%-38% of all maternal deaths [11]. Clinical imaging of preeclampsia demonstrated brain edema and cerebrovascular hemorrhage [12-15], the cerebral circulation has been suggested to play a central role in neurological complications of preeclampsia, and thus understanding how pregnancy and preeclampsia affect the cerebrovasculature is of interest [15,16]. Therefore, preeclampsia is thought to be a hypertensive encephalopathy. Investigating how cerebral hemodynamics change during pregnancy including microvascular structure, may provide important mechanisms underlying preeclampsia. Understanding how normal pregnancy affects the cerebral vascular environment is important for neurological conditions such as preeclampsia. Therefore, the present study aimed to investigate dynamic changes in the blood flow and vessel diameter from prepregnancy to late pregnancy by in vivo two-photon imaging.

Materials and Methods

Animals

A total of 12 female adult c57 mice (weight 20-22 g, 10-12 weeks) were obtained from experimental animal center of southern medical university (Guangzhou, Guangdong, China), the mice were acclimatized to the laboratory conditions for 1 week prior to the start of the experiments. Each mouse in all the pregnancy groups was separately mated overnight. Day 0 of pregnancy was defined as the day when a sperm plug was found in a vaginal smear [17]. Animals were grouped as following: the non-pregnant group (n=3), early-pregnant group (n=3, day=3), middle-pregnant group (n=3, day=10), late-pregnant group (n=3, day=17). Experimental protocols were approved by the Committee on the Ethics of Animal Experiments of Guangzhou Medical University (Permit Number: 2012-50) in accordance with the NIH Guidelines (NIH Publications No. 8023, revised 1978) for the Care and Use of Laboratory Animals. All efforts were made to minimize the number of animals used as well as their suffering.

Thinned-skull window preparation for in vivo imaging

To make a cranial window for imaging, mice were anaesthetized with 1.5% tribromoethanol (20 ml/kg, i.p.), the most part of brain skin was removed to expose the skull. The position of window was 2.0 mm posterior to the bregma and 2.0 mm lateral from the middle line. At the beginning, a micro drill (Ideal, Cat® SN3150) was used to thin a 1-2 mm diameter circular skull region, after removing the majority of the spongy bone, remaining concentric cavities within the bone can usually be seen under the dissecting microscope, indicating that drilling is approaching the internal compact bone layer. At this stage, skull thickness should still be more than 50 μm. Then a 10° surgical blade was used to slowly and carefully thin the skull until the blood vessels on the cerebral cortex were clearly visible under a light microscope (a ~20 μm preparation). During the whole thinning process, 0.9% physiological saline was added to the skull surface from time to time to reduce heat. After the thinned skull became thoroughly dry, a small drop of thin glue (TED PELLA, INC. Cat®1003) was applied and a small piece of cover glass (about 1-1.5 × 1-1.5 mm size) was glued onto the thinned skull. The remaining area of the skull was covered with a layer of the thin glue. The mice were allowed to recover for least 1 day before imaging.

Two-photon imaging

The mice were anaesthetized with 1.5% tribromoethanol (20 ml/kg, i.p.) and received an intraperitoneal injection of 0.1 ml Rhodamin B sothiocyanate-dextran solution (70 KDa, 3 mg/ml in distill water) at least 15 min before imaging to label the blood plasma. The mice were placed on a head holder (Narishigie, Cat®SG-4N) and the head position was adjusted so that the cortical surface was in a horizontal position to the microscope objective. The mice were placed onto of a healing pad to maintain the body temperature at 37°C, and imaging was made with a two photon microscopy. We used 870 nm excitation wavelength (Maitai DeepSee, 10 RP52-2, Newport, USA). A 20X objective lens was used to image individual blood vessels and take line scan images along with the central axis of a vessel at 3X optical zoom. Each session of line scan consisted 5000 lines. The line scan parameters were automatically recorded in the ZEN software.

Analysis of blood flow and vessel diameter

A program in MatLab written by Dr Xiao Ming Jin and his student was used for calculating the velocity of Red Blood Cells (RBC) in cerebral blood vessels according to the line scan parameters. The vessel diameter was measured by NIH Image J.

Statistical analysis

A one-way Analysis of Variance (ANOVA) was used to compare the average values between the different groups, which was followed by an appropriate Least Significant Difference (LSD) post hoc test (homogeneity test of variance is not neat) or else Dunnett's test (homogeneity test of variance is not neat). Values were presented as Mean ± SE and statistical significance was denoted as P<0.05.
Results

The morphologies, blood flow signals and blood flow velocity of Rhodamin stained microvessels

Figures 1A and 1B showed the morphology and the fragment of an arteriovenous vessel and a capillary. The blood flow signals of the arteriovenous vessel (Figure 1A-1) and the capillary (Figure 1B-1) can be clearly seen; especially single black line can be seen in the capillary blood flow signal (Figure 1B-1), as we all know the capillary allows only one blood cell to go across, single black line indicates one blood cell. The velocity (Figure 1A-2) of arteriovenous vessel seemed much higher than that of the capillary (Figure 1B-2). Scale bar=50 μm.

Changes in blood flow velocity from non-pregnancy to late-pregnancy

300 arteriovenous vessels or capillaries in each group were selected at random for the measurement of velocity. Quantitative analysis demonstrated that the blood velocity of arteriovenous vessels significantly increased in the middle-pregnancy group (4.54 ± 0.44 mm/s) compared to the values in the non-pregnancy (4.14 ± 0.51 mm/s) and early-pregnancy (4.21 ± 0.47 mm/s) groups (P<0.05, Table 1); the blood velocity of arteriovenous vessels in the late-pregnancy group (5.44 ± 0.70 mm/s) was significantly higher than in the non-pregnancy, early-pregnancy and middle-pregnancy groups (P<0.05), Figure 2A. The velocity of capillaries in the late-pregnancy group (0.48 ± 0.07 mm/s) also significantly increased compared to the values in the non-pregnancy (0.33 ± 0.06 mm/s), early-pregnancy (0.34 ± 0.03 mm/s) and middle-pregnancy (0.37 ± 0.07 mm/s) groups (P<0.05), Figure 2B.

Changes in morphology of vessels from non-pregnancy to late-pregnancy

From non-pregnancy to early-pregnancy, we didn’t observe vasodilation (Figures 3A and 3B and 3E and 3F). Obvious vasodilation of arteriovenous vessels (Figures 3C and 3D) can be seen in late-pregnancy. Though the diameter of capillaries is originally smaller, slight vasodilation still can be seen (Figures 3G and 3H). Figures 3A-3D: Scale bar=50 μm. Figures 3E-3H: Scale bar=20 μm.

Changes in vessel diameter from non-pregnancy to late-pregnancy

300 arteriovenous vessels and 300 capillaries in each group were selected at random for the measurement of diameter. The Image J was used to measure the diameter by using the parameters which were corresponding with the morphological pictures. Statistical analysis revealed that the diameter of arteriovenous vessel significantly increased in the middle-pregnancy (32.36 ± 1.82 μm) and late-pregnancy (35.88 ± 3.64 μm) groups than in the non-pregnancy (29.05 ± 3.13 μm), early-pregnancy (30.29 ± 2.18 μm) groups (P<0.05, Table 2); the diameter of arteriovenous vessel in the late-pregnancy

Evolution of cerebrovascular changes in mouse cortex during normal pregnancy by in vivo two-photon imaging
group also became bigger compared to that in the middle-pregnancy group (P<0.05), Figure 4A. The diameter of capillary also increased significantly middle-pregnancy (5.02 ± 0.67 μm) and late-pregnancy (5.55 ± 0.82 μm) groups than in the non-pregnancy (4.01 ± 0.62 μm), early-pregnancy (4.31 ± 0.64 μm) groups (P<0.05), Figure 4B.

Discussion
This study showed that the blood flow velocity of arteriovenous vessels and capillaries in late-pregnancy and middle-pregnancy increased by 8%-30% in comparison with that in non-pregnancy and early-pregnancy; the vessel diameter in late-pregnancy and middle-pregnancy increased by 7%-24% compared to that in non-pregnancy and early-pregnancy. The changing tendency of blood flow velocity is consistent with that of vessel diameters. We concluded that maternal cerebral blood flow is gradually increasing during normal pregnancy; the morphological changes of vessels might be related to the blood flow changes.

During pregnancy, there is a large increase in cardiac output and plasma volume, the uterine blood flow in late-gestation increased 10-fold over the non-pregnancy; but the brain is enclosed in a rigid skull, dramatic increase in cerebral blood flow could result in detrimentally increased intracranial pressure that can cause serious neurological symptoms, brain edema, cerebral hemorrhage and even death. So the magnitude by which cerebral blood flow change needs to be assessed accurately.

Using dual-beam angle-independent digital Doppler ultrasound, Ori et al. found that a maximum increase of 20% in the maternal blood flow of the Internal Carotid Artery (ICA) during the course of pregnancy compared to the nonpregnant baseline, such increase is associated with moderate increase in ICA diameter [18]. Another study showed 8.1% increase in cerebral hemispheric blood flow during the first trimester by using Single-Photon Emission Computerized Tomography (SPECT) before and 6 weeks after pregnancy termination [19]. Our results are in accordance with these reports.

The adaptation of cerebral circulation during pregnancy is related to the vasodilatory effects of Vascular Endothelial Growth Factor (VEGF), estrogens and other circulating factors [18,20]. Rusterholz and Aagaard-Tillery et al. proposed that hormones production including growth factors, cytokines increases significantly throughout pregnancy, reaching a peak in late gestation [2,21]. VEGF is one of the most important growth factors for successful pregnancy, it has been recognized as having a role in angiogenesis, vascular growth, endothelial cell survival, and vasodilation through complex interaction of VEGF with its receptors [22,23]. In the uterine circulation and cerebral veins, VEGF expression is higher than the nonpregnant condition [24,25]. Estrogen (ER) may also promote vasodilation during pregnancy; ER expression in endothelial and vascular smooth muscle layers of the aorta and mesenteric artery increased during pregnancy [26]. The related mechanisms for ER mediated vasodilation may be through inhibition of Ca$^{2+}$ entry into vascular smooth muscle and increasing NO formation by increasing Ca$^{2+}$ activated endothelial NOS expression or by increasing substrate availability for NO formation [27,28].

Our previous studies has proposed that normal pregnancy lowered seizure threshold [29,30], one possible mechanism was that mild systemic inflammation might cause neuroinflammation which could increase neuronal excitability during pregnancy [1,29-32]. Another possible mechanism may be related to cerebral blood flow autoregulation breakdown. When the extent by which blood flow changed was beyond the scope of brain autoregulation during pregnancy, brain blood barrier would be disrupted and then caused cerebral edema formation. Clinical reports have shown that 93% of eclamptic women had cerebral vasogenic edema in diffusion-weighted MRI studies [33]. The present study demonstrated that in late-pregnancy the blood velocity increased by 8%-30% compared with that in non-pregnancy. Such changed magnitude of blood flow during pregnancy can served as a reference to assess whether the brain blood barrier is intact and the possibility of eclampsia-like seizures.

Different from clinical imaging techniques, the application of two photon imaging in this study helped us to obtain high-resolution images of microvasculature of the cortex in vivo, the diameter and blood flow velocity of each microvessel can be exactly measured. The present findings also provided some basis in investigating how microvascular environment change in preeclampsia and eclampsia which is a leading cause of maternal morbidity and morality worldwide [34]. However, the CBF changes in preeclampsia and eclampsia is not clear. Since molecular transport across the blood-brain barrier mainly occurs in the microvasculature (especially the capillaries), all the vessels play an important role in the metabolism, energetics, and functionality of the cortex [35]. Further studies
Evolution of cerebrovascular changes in mouse cortex during normal pregnancy by in vivo two-photon imaging

needed to be performed to investigate changes in the diameter and blood flow velocity of each microvessel under pathological conditions such as preeclampsia and eclampsia.

Table 1. Blood flow velocity during pregnancy.

<table>
<thead>
<tr>
<th>Group (n)</th>
<th>Number (n)</th>
<th>Non-pregnancy (mm/s)</th>
<th>Early-pregnancy (mm/s)</th>
<th>Middle-pregnancy (mm/s)</th>
<th>Late-pregnancy (mm/s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Arteriovenous</td>
<td>1200</td>
<td>4.14 ± 0.33</td>
<td>4.21 ± 0.41</td>
<td>4.54 ± 0.29</td>
<td>5.44 ± 0.58#</td>
</tr>
<tr>
<td>Capillary vessel</td>
<td>1200</td>
<td>0.33 ± 0.12</td>
<td>0.34 ± 0.09</td>
<td>0.37 ± 0.07</td>
<td>0.48 ± 0.10#</td>
</tr>
</tbody>
</table>

p<0.05, compared with the Non-pregnancy group and Early-pregnancy group

Table 2. Vessel diameter during pregnancy (um).

<table>
<thead>
<tr>
<th>Group (n)</th>
<th>Number (n)</th>
<th>Non-pregnancy (um)</th>
<th>Early-pregnancy (um)</th>
<th>Middle-pregnancy (um)</th>
<th>Late-pregnancy (um)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Arteriovenous</td>
<td>1200</td>
<td>29.05 ± 2.96</td>
<td>30.29 ± 3.41</td>
<td>32.35 ± 2.17</td>
<td>35.88 ± 4.58#</td>
</tr>
<tr>
<td>Capillary vessel</td>
<td>1200</td>
<td>4.01 ± 1.41</td>
<td>4.31 ± 1.62</td>
<td>5.02 ± 1.68</td>
<td>5.55 ± 1.95#</td>
</tr>
</tbody>
</table>

p<0.05, compared with the Non-pregnancy group and Early-pregnancy group

Conflict of Interest

The authors declare that there is no conflict of interest.

Acknowledgments

This study was supported by the National Natural Sciences Foundation of China (9681170594), the Guangzhou Science and Technology Project (2016A020218002).

References


*Correspondence to*

Huishu Liu
Department of Obstetrics
Guangzhou Women and Children’s Medical Center
PR China