

## Cloning and bioinformatics analysis of a glutamate decarboxylase from *Lactobacillus plantarum* LpS2.

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### Abstract

A *Lactobacillus plantarum* (LpS2) that was found to produce a large amount of  $\gamma$ -aminobutyric acid (GABA) was isolated from yellow serofluid, a bean curd byproduct in China. a) Glutamate decarboxylase (GAD) gene from *Lactobacillus plantarum* LpS2 was Cloned and amplified by PCR technology. Then, we predicted its amino acid composition, physicochemical property, signal peptide and advanced structure by on-line analysis tools and software. b) last analysis results showed that the amplification of gene nucleotide sequences have a very high homology with lpgad B reported in NCBI gene sequences, and the deduced amino acid sequences homologies were as high as 99%. The length of gene sequence was 1363 bp and coded 453 Amino acids. GAD relative molecular weight of theoretical prediction and isoelectric point were 51521.8u and 5.39 respectively. This protein was hydrophilic protein, and there was no transmembrane region. The analysis results laid a theoretical basis for construction of genetic engineering bacteria.

**Keywords:** Glutamate decarboxylase; *Lactobacillus plantarum*; Bioinformatics Analysis  $\gamma$ -aminobutyric acid.

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### Introduction

$\gamma$ -Aminobutyric acid (GABA) is a non-protein amino acid that is widely distributed in nature and plays an important role as the major inhibitory neurotransmitter in the mammalian central nervous system [1]. It has such physiological functions as the induction of hypotensive, diuretic, and tranquilizing effects. It has also been implicated in the regulation of several neurological disorders, including Parkinson's disease, Alzheimer's disease, and Huntington's chorea. Due to its physiological functions, the increasing commercial demand for GABA has prompted studies on the development of Health food and pharmaceutical fields [2-5].

The current production of GABA methods mainly include chemical method and biological synthesis. Chemical synthesis method has the following disadvantages: chemical residues, poor safety, high cost. Microbial fermentation is a method of preparation of GABA with high efficiency and low cost. Numerous studies on GABA production from various microorganisms have been reported [6-9].

GABA is primarily produced by the irreversible  $\alpha$ -decarboxylation of acidic glutamate in a reaction catalyzed by glutamate decarboxylase (GAD). Glutamate decarboxylase (GAD) is an essential enzyme widely distributed in nature from microorganisms to plants and animals. It is the key enzyme for the biosynthesis of GABA. So the studies of scientists are still focus on the mechanism of GAD now. To

find a high GAD activity of strain will become an important study for researchers [10-11].

In this study, we isolated a new GABA-producing microorganism from yellow serofluid, a bean curd byproduct of production in China. This bacterial strain, LpS2, was identified as *Lactobacillus plantarum*. In addition, we amplified the gene of GAD from the LpS2. Then, we predicted its amino acid composition, physicochemical property, signal peptide and advanced structure by bioinformatics method. So that we can more fully understand the characteristics of GAD gene from *Lactobacillus plantarum* LpS2, and build a foundation for the subsequent genetic engineering bacteria. Eventually we will be able to use genetic engineering bacteria to produce GABA food and drugs by microbial fermentation method.

### Materials and Methods

#### *Strains, media and growth conditions*

The *Lactobacillus plantarum* LpS2 that was found to produce a higher amount of  $\gamma$ -aminobutyric acid (GABA) was isolated from yellow serofluid. Centrifugal type column bacterial genome DNA extraction kit DP302-2 was produced by tian gen biological engineering Ltd. Centrifugal type column DNA gel recovery kits (DP209-02) was produced by tian gen biological

engineering Ltd. Trans taq hifi DNA Polymerase was produced by Shanghai treasure biological technology Ltd.

MRS liquid medium consists of 1% peptone, 0.5% yeast powder, 1% beef powder, 0.5% anhydrous sodium acetate, 0.2% citric acid diammonium hydrogen, 1% glucose, 0.05% MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.02% MnSO<sub>4</sub>, 0.2% K<sub>2</sub>HPO<sub>4</sub>, 0.1% Tween 80, pH 6.8, Liquid medium were sterilized at 121°C for 15 min before use.

### Cloning and DNA sequencing

According to the inoculation quantity of 2%, the strains LpS2 were inoculated into MRS liquid medium, incubated at 37°C for 48 h. The 1.5 ml activated bacterium solution was placed in the 2 ml centrifuge tube. The cells of LpS2 were collected by centrifuging at 10000 rpm for 1 min. The supernatant was discarded. 180 µl lysozyme (20mg/ml) were added into cells for wall-breaking treatments and constant temperature at 37°C for 30 min. Then 20 µl Proteinase K solution (20mg/ml) was added into solution for mixing.

Using the kit method for DNA extraction, solution were collected to the centrifugal tube and analyzed by 1% agarose gel electrophoresis (150 v, 100 mA) for 20 min. Then the gel of cells genomic DNA was recycled by kit method. The collected DNA was dissolved into 30µL ddH<sub>2</sub>O.

*Lactobacillus plantarum* LpS2 GAD gene was amplified by kit method. GadB-f(forward): 5'-GTAGAATTCATGGCAATGTTATACGGTAAACAC-3' and GadB-r(reverse): 5'-CTAGCGGCCGCGTGTGTGAATCCGTATTTCTTAG-3' were designed by reported *Lactobacillus plantarum* GAD gene sequence (Primer design was completed by sangon biological engineering co.). The total DNA was taken as a template for amplification (Table 1). By the method of kit, *Lactobacillus plantarum* GAD gene PCR products were recycled.

**Table 1:** The system of reaction.

genomeDNA	1 µl
GadB-F	1 µl
GadB-R	1 µl
10×PCR Buffer	5 µl
dNTP mix (2.5 mmol/L)	4 µl
Hifi (5 U/µL)	1 µl
ddH <sub>2</sub> O	37 µl
total volume	50 µl

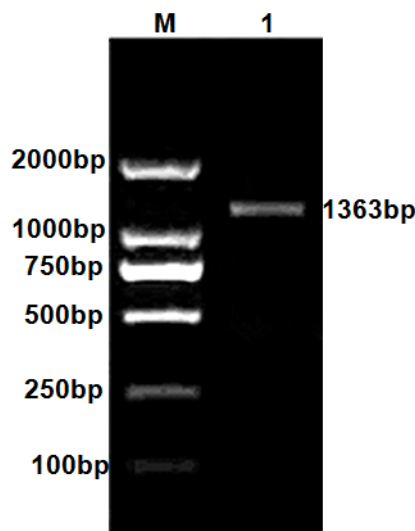
The GAD gene amplification products sent to the sangon biological engineering (Shanghai) LTD for Sequencing. The sequencing results were analysed through Exploring-Local Vector NTI Database software and translated nucleotide sequence homology. Then the physical and chemical properties of protein, feeling of water analysis, protein signal peptide

prediction, N-glycosylation sites prediction and protein structure domain were analysed respectively [12-15].

## Results and Discussion

### PCR amplification of *Lactobacillus plantarum* GAD gene

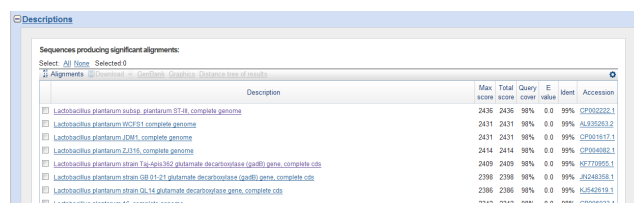
Strain GAD gene was amplified by PCR, and then amplification products were analyzed by agarose gel electrophoresis. The result of PCR amplification of *Lactobacillus plantarum* GAD gene was shown in Figure 1.



**Figure 1:** Result of PCR amplification of *Lactobacillus* GAD gene. M: DL2000 Marker; 1: PCR products of LpS2.

### Sequencing result of *Lactobacillus plantarum* GAD gene

We obtained two gene sequences by sequencing. The two gene sequences were joined together into a gene segment by software Exploring Local Vector NTI Database, which length was 1363 bp and encode 453 amino acids. By landing on NCBI (National Center of Biotechnology Information) and Using Blast program, gene segment was searched for nucleotide homology. The result was shown in Figure 2.



**Figure 2:** Blast results of *Lactobacillus* GAD gene nucleotide sequence.

Blast analysis results showed that the amplification of gene nucleotide sequences had a very high homology with lpgadB reported in NCBI gene sequences (GenBank Accession No. KF770955.1). The deduced amino acid sequences homologies was as high as 99%. Through logged in website (<http://>

web.expasy.org/translate/) to translation, the result showed there were a total of 453 amino acids (Figure 3).

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1 aaitcatgacagctgagatgacgtgaccagtcttgggtcgcctctcgaacaacatgatc
F M Q A E I L G P V F G A P S E Q H D L
61 ttccaaagtatcggtaaccaagcattcattatccctcgaagaagccgatcgctttagttc
P K Y R L P K H S L S P R E A D R L V R
121 gtgatgaattattagatgaggaacttcaacgactgaaactgttggcagactct
D E L L D E G N S R L N L A T F C Q T Y
181 atatggaaccggaagccgttgatgaaaggatacctgctcgaagcaatgcacgaca
M E P E A V E L M K D T L A K N A I D K
241 aatctgagtaaccocccacggccgagattgaaaacgggtgtgaaacattattgcacatc
S E Y P R T A E I E N R C V N I I A N L
301 tgtgacagcaactgatgacgaacactttaaggtaoctctcgaattggctcctcgaag
W H A P D D E H F T G T S T I G S S E A
361 ctgtatgttagcgttttagcaatgaaattcgcctggcgtaaacgctcaagcggcgg
C M L G G L A M K F A W R K R A Q A A G
421 gtttagatcgaatgcccacgacctaaccctgttatttcggctggctatcaagtttct
L D L N A H R P N L V I S A G Y Q V C W
481 gggaaaagtgttctgctactggacgttgacatgcaactgctccaatggatgagcaac
E K F C V Y W D V D M H V V P M D E Q H
541 acatggcccttgacgttaaccacgtcttagactaactgacgaaacaaattggtatcg
M A L D V N H V L D Y V D E Y T I G I V
601 tggatcagcggcaccatcattaccggcgaatgacgacactagccgcaactcgaagg
G I M G I T Y T G Q Y D D L A A L D K V
661 tcttactactacaatcagcactccaaattaaccgctcaactcagttgacgcaag
V T H Y N H Q H P K L P V Y I H V D A A
721 cgtcaggtgctctataccocattattgagcgcacacacactcggacttcgggtgg
S G G F Y T P F I E P Q L I W D F R L A
781 ctaacgtctgttcgatcaacgctcgggacacaagtaagggttttagttatccgggtcg
N V V S I N A S G H K Y G L V Y P G V G
841 gctgggtctgttgctgctgacgtttttaccggcgaatgcttcaagtttagtt
W V V W R D R Q F L P P E L V F K V S Y
901 atttaggtgggaggttccgcaaatggcgcacactctcacaatgacgacccagctca
L G G E L P T M A I N F S H S A A Q L I
961 ttggacaactataatttcttctgtgtagcgggtaaccgagattcaaacaa
G Q Y Y N F I R F G M D G Y R E I Q T K
1021 agactcagatgttgcocctacctgacagcgcctctggataaattggtgagtttaag
T H D V A R Y L A A A L D K V G E F K M
1081 tgatcaatacggacaccaactccccctgattgttaccactagcccgcgcaagatc
I N N G H Q L P L I C Y Q L A P R E D R
1141 gtgaatggaactttatgattatcgatcgcctattaatgaaacgggtggcaggtacaa
E W T L Y D L S D R L L M N G W Q V P T
1201 cgtatccttactcctaatctggaacaacaaatgcacccaaagatcgtcttccggctg
Y P L P A N L E Q Q V I Q R I V V R A D
1261 actttggcatgaatggcccacgatttcgatgacgacacaaaggctgtocagatc
F G M N M A H D F M D D L T K A V H D L
1321 taaaccaagcccactgtcttcatcagaccccccccccc1363
N Q A H I V F I M T P A P
    
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Figure 3: Translation results of *Lactobacillus GAD* gene nucleotide sequence.

**Bioinformatics analysis of *Lactobacillus plantarum* GAD**

By logging on website (<http://web.expasy.org/protparam/>) for online analysis, the formula of protein (GAD) was C2326H3541N619O663S23 and the proteins were composed of 7172 atoms. The molecular weight was 51.5 kDa and an estimated isoelectric point (pI) was 5.39. Amino acid compositions of protein (GAD) were shown in Table 2.

**Protein hydrophilic and transmembrane region analysis of *Lactobacillus plantarum* GAD**

By logging on website (<http://web.expasy.org/protscale/>) and website ([http://www.ch.embnet.org/software/TMPRED\\_form.html](http://www.ch.embnet.org/software/TMPRED_form.html)) for protein hydrophilic and transmembrane region online analysis, ProtScale (a programme of ExPASy) predicted that hydrophobic amino acids GRAVY were greater than zero, and the hydrophilic amino acids GRAVY were less than zero. GRAVY of protein (GAD) was

0.179. The result indicated that the protein (GAD) was hydrophilic protein (Figures 4 and 5).

Table2: Amino acid composition of *Lactobacillus GAD* gene.

Amino Acid	Quantity	Percentage %
AlaA	39	8.6
ArgR	21	4.6
AsnN	20	4.4
AspD	32	7.1
CysC	6	1.3
GlnQ	20	4.4
GluE	24	5.3
GlyG	28	6.2
HisH	19	4.2
IleI	25	5.5
LeuL	43	9.5
LysK	16	3.5
MetM	17	3.8
PheF	18	4.0
ProP	25	5.5
SerS	16	3.5
ThrT	19	4.2
TrpW	9	2.0
TyrY	22	4.9
ValV	34	7.5
PylO	0	0.0
SecU	0	0.0
B	0	0.0
Z	0	0.0
X	0	0.0

**Signal peptide prediction of *Lactobacillus plantarum* GAD**

By logging on website (<http://www.cbs.dtu.dk/services/SignalP/>) for online analysis, the results of prediction were shown in Figure 6. By the value of C,S and Y value could be judged that there was no signal peptide in this protein. After the synthesis of GAD in the cytoplasm, protein was not transported. The results showed that GAD protein was not secreted proteins or some cytoplasmic matrix synthesis proteins, which was transported to organelles and played a role in there.

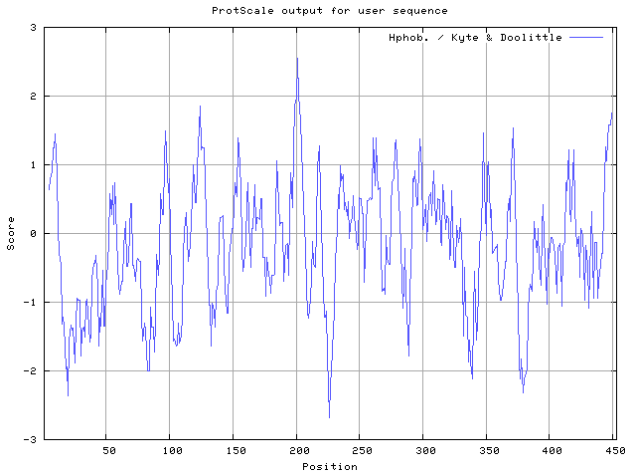


Figure 4: Analysis of Hydrophilic for protein.

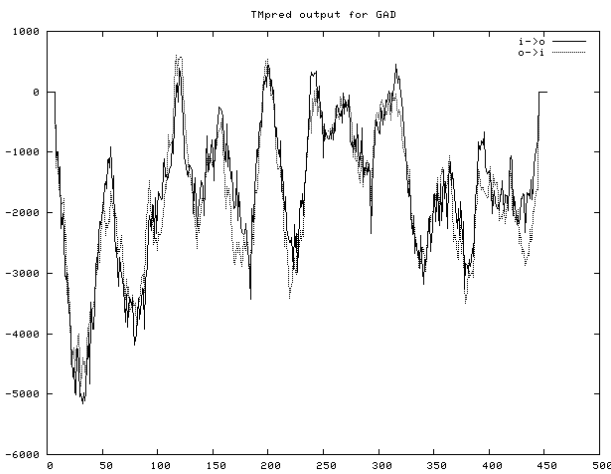


Figure 5: Analysis of transmembrane region for protein.

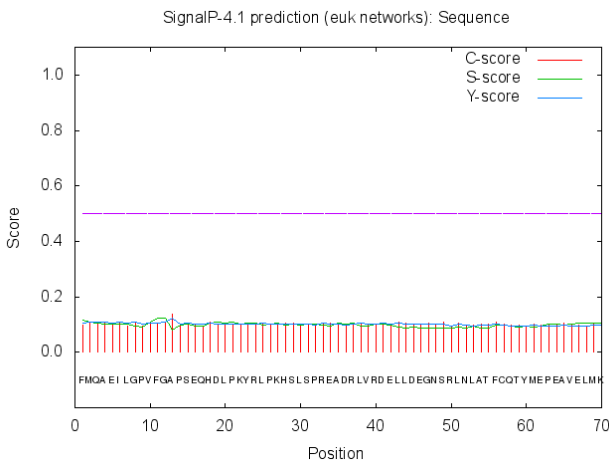
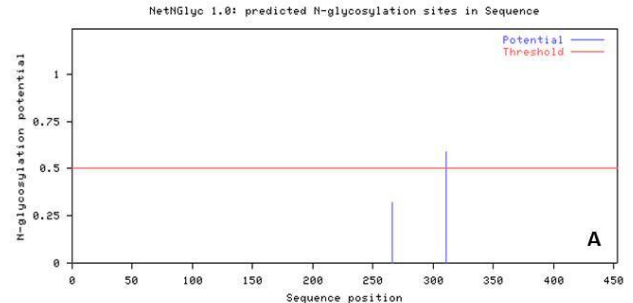


Figure 6: Prediction of signal peptide for protein.

### N-glycosylation analysis of *Lactobacillus plantarum* GAD

N-glycosylation is an important step of protein modification after translation. In addition, N-glycosylation has an influence

on cell biology behavior of protein such as antigenicity, active, folding, transport, positioning and stability. By logging on Website (<http://cbs.dtu.dk/services/NetNGlyc>) to analysis and predict N - glycosylation sites, the result displayed that the N-glycosylation sites had 2 points (Figure 7).



SignalP output is explained at <http://www.cbs.dtu.dk/services/SignalP/output.html>

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Name: Sequence Length: 453
FRKAEILGPPGAPSEQHDLPKYRPFRESLSPREADRVRDELLDEGMSRLMLATPCQTYHEPEAVELMKD LAKNAIDK 80
SEYPTAEIENRCVHIIANLWHAPDDEHF TGTSTIGSSEACHLGLAMKFAWRKQAAGLDLNAHRPVLISAGYQCVW 160
KEPCYTDVDRHVPDDEQHMLDVHVLDTYDEYITIGVINGITITGQYDILAAALDKVYIYHHRQPKLPTVTHVDA 240
SGGFTTFPIEIQIIDFRLAVYSIRASGSEKGLVYPCGCVYFRDRQLPHELYFSTYIGGLGELPTALHFSHSAAGL 320
GQTYHFIRFGNDGYSIEIQTKHDAVARYLAAALDKVGFEMINRCHOLPLICVGLAPREDREWTLTDLSDRLIMRCWQVPT 400
YPLPAHLEQQVIQIYVYRADFCNHHADFDHDLTKAVHDLNQAHIVFINTPAP 480
..... 80
..... 160
..... 240
..... 320
..... 400
..... 480

```

(Threshold=0.5)

SeqName	Position	Potential	Jury agreement	N-Glyc result
Sequence	266 HASC	0.3173	(9/9)	---
Sequence	311 NFSH	0.5860	(7/9)	+

Figure 7: Prediction and analysis of N- glycosylation sites for protein.

### Structure domain analysis of *Lactobacillus plantarum* GAD

By logging on website (<http://www.ncbi.nlm.nih.gov/Structure/cdd/>) and using the connection in the NCBI CDD structure domain analysis, the result was shown as follow (Figure 8). *Lactobacillus plantarum* GAD was in the amino acid sequence 9~441 position for structural function domain. GAD gene belonged to aspartate amino transferase (AAT) superfamily (type I) pyridoxal phosphate (PLP) dependence enzymes. The comparison test results with family members (TIGR01788) were shown in Figure 9.

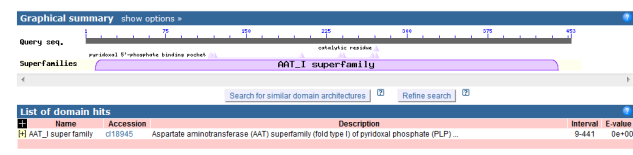


Figure 8: Analysis of domain for protein.

### Secondary structure prediction of *Lactobacillus plantarum* GAD

Through logging on website ([http://npsa-pbil.ibcp.fr/cgi-bin/npsa\\_automat.pl?Page=npsa\\_sopm](http://npsa-pbil.ibcp.fr/cgi-bin/npsa_automat.pl?Page=npsa_sopm)). HTML and <http://www.cbs.dtu.dk/services/TMHMM-2.0/>, the secondary structure and transmembrane helical of GAD were analysed. The result was shown in Figure 10 and Figure 11. As shown in Figure 10, we used the SOPM online to predict. The GAD secondary structure was composed of 43.27% alpha helix (Hh),

19.65% extension chain (Ee), 29.14% random curl(Cc) and 7.94% beta Angle (Tt). As shown in Figure 11, the protein transmembrane helical was no signal. The result indicated that the protein was not secreted proteins, and there was no transport across the membrane.

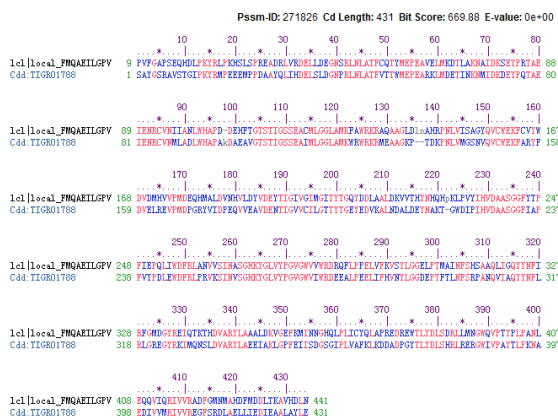


Figure 9: Comparison chart of family members TIGR01788.

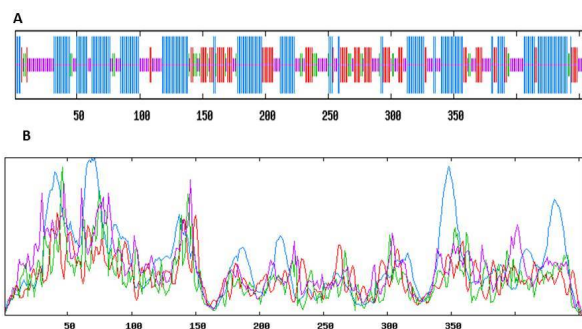


Figure 10: Prediction of Secondary Structure of protein.

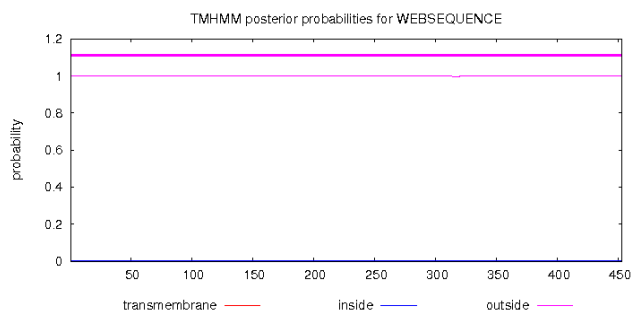


Figure 11: Analysis of transmembrane helix of protein.

### Tertiary structure prediction of *Lactobacillus plantarum* GAD

Using homology modeling method, we logged in web site (<http://swissmodel.expasy.org/>) for predicting the tertiary

Table 3: Modeling contrast of *Lactobacillus* GAD gene.

structure of GAD gene sequence. Analysis results were shown in Figures 12 and 13. We took F chain of 1xey.1. As a template, and shaped residue cover wide range of 3-432 for modeling Blast by X-ray method. The consistency of both sequences was 46.01%. Low poly state was homologous six polymers. Resolution was 2.5A. Requence similarity was 0.41. Coverage rate was 94%. GMQE value was 0.77.QMEAN4 value was -3.02.

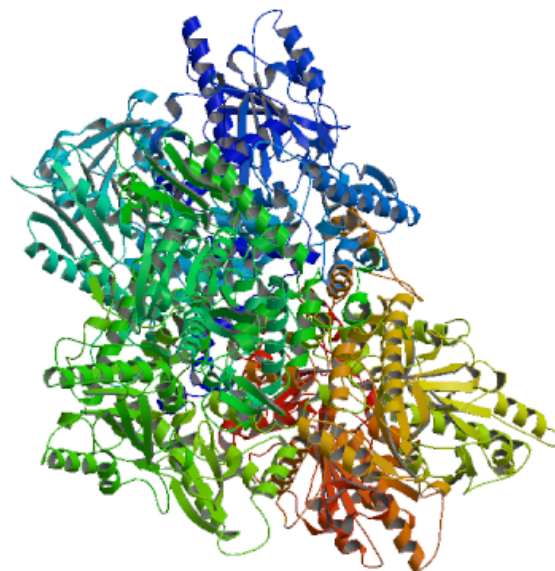


Figure 12: Three-dimensional structure of protein.

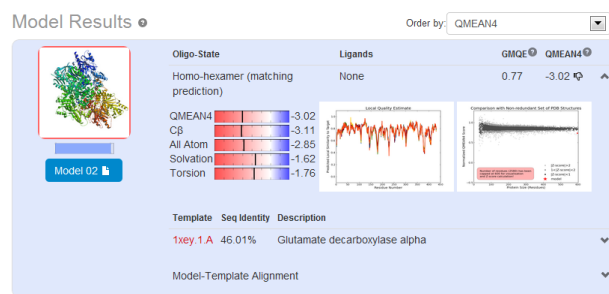


Figure 13: Report of transmembrane helix of protein.

Comparison of different template on protein modeling (table 3), we took 3fz6.1.A1pmm.1.A1xey.1.A\_2dkg.1.A and 3hbx.1.A as template for modeling. The coverage could reach more than 92%, and the consistency could reached more than 42%. The above showed that the gene belonged to the GAD gene.

Template	Seq Identity	Oligo-state	Found by	Method	Resolution	Seq Similarity	Coverage	Description
3fz6.1.A	45.21	homo-hexamer	HHblits	X-ray	2.82 Å	0.43	0.97	Glutamate decarboxylase beta
1pmm.1.A	45.21	homo-hexamer	HHblits	X-ray	2.00 Å	0.43	0.97	Glutamate decarboxylase beta
1xey.1.A	45.08	homo-hexamer	HHblits	X-ray	2.05 Å	0.42	0.96	Glutamate decarboxylase alpha
2dgk.1.A	45.16	homo-hexamer	HHblits	X-ray	1.90 Å	0.42	0.96	Glutamate decarboxylase beta
3fz6.1.A	46.14	homo-hexamer	BLAST	X-ray	2.82 Å	0.43	0.94	Glutamate decarboxylase beta
1pmm.1.A	46.14	homo-hexamer	BLAST	X-ray	2.00 Å	0.43	0.94	Glutamate decarboxylase beta
1xey.1.A	46.01	homo-hexamer	BLAST	X-ray	2.05 Å	0.43	0.94	Glutamate decarboxylase alpha
3hbx.1.A	40.46	homo-hexamer	HHblits	X-ray	2.67 Å	0.41	0.96	Glutamate decarboxylase 1
2dgk.1.A	46.65	homo-hexamer	BLAST	X-ray	1.90 Å	0.43	0.92	Glutamate decarboxylase beta
3hbx.1.A	42.11	homo-hexamer	BLAST	X-ray	2.67 Å	0.42	0.92	Glutamate decarboxylase 1

## Conclusion

In this study, a new GABA producing microorganism, *Lactobacillus plantarum* LpS2 was isolated from yellow serofluid. A full-length GAD gene from LpS2 was cloned. The amino acid sequence of GAD was analysed. The length of GAD gene sequence was 1363 bp and coded 453 Amino acids. GAD relative molecular weight of theoretical prediction and isoelectric point were 51521.8u and 5.39 respectively. This protein was hydrophilic protein, and there was no transmembrane region. There were two N-glycosylation sites. Structure and analysis showed that the protein belonged to the AAT super family (fold type I) pyridoxal phosphate (PLP) dependence enzyme. Secondary structure was composed of 43.27% Alpha helix, 19.65% extension chain, 29.14% random curl, 7.94% beta Angle. Transmembrane helical was no signal, and there was no transmembrane transport. The analysis of tertiary structure showed that 3fz6.1.A-1 pmm.1.A-1xey.1.A-2dgk.1.A and 3hbx.1. A were taken as template for modeling, and the coverage could reach more than 92%, and the consistency could reached more than 42%.

This research adopted the bioinformatics analysis method, which was considered to be authoritative in the biological field. This result was very important for the expression, separation and purification of glutamate decarboxylase in the following work. In addition, it is very meaningful for producing GABA food and drug by genetic engineering strains.

## Acknowledgment

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## References

1. Manyam BV, Katz L, Hare TA, Kaniefski K, Tremblay RD. *Ann Neurol*, 1981; 10: 35-37.
2. Hayakawa K, Kimura M, Kasaha K, Matsumoto K, Sansawa H, Yamori Y. Effect of a gamma-aminobutyric acid-enriched dairy product on the blood pressure of spontaneously hypertensive and normotensive Wistar-Kyoto rats. *Br J Nutr* 2004; 92: 411-417.
3. Inoue K, Shirai T, Ochiai H, Kasao M, Hayakawa K, Kimura M, Sansawa H. Blood-pressure-lowering effect of a novel fermented milk containing gamma-aminobutyric acid (GABA) in mild hypertensives. *Eur J Clin Nutr* 57: 490-495.
4. Jakobs C, Jaeken J, Gibson KM. Inherited disorders of GABA metabolism. *J Inherit Metab Dis* 1993; 16: 704-715.
5. Wong CG, Bottiglieri T, Snead OC. GABA, gamma-hydroxybutyric acid and neurological disease. *Ann Neurol* 2003; 54: S3-12.
6. Nomura M, Nakajima I, Fujita Y, Kobayashi M, Kimoto H, Suzuki I, Aso H. *Lactococcus lactis* contains only one glutamate decarboxylase gene. *Microbiology* 1999; 145: 1375-1380.
7. Park KB, Oh SH. Cloning, sequencing and expression of a novel glutamate decarboxylase gene from a newly isolated lactic acid bacterium, *Lactobacillus brevis* OPK-3. *Bioresource Technology* 2007; 98: 312-319.
8. Park KB, Heung OS. Cloning and expression of a full-length glutamate decarboxylase gene from *Lactobacillus plantarum*. *Journal of Food Science and Nutrition* 2004; 9: 324-329.

9. Kim SH, Shin BH, Kim YH. Cloning and expression of a full-length glutamate decarboxylase gene from *Lactobacillus brevis* BH2. *Biotechnology and Bioprocess Engineering* 2007; 12: 707-712.
10. Naidu AS, Bidlack WR, Clemens RA. Probiotic spectra of lactic acid bacteria (LAB). *Crit Rev Food Sci Nutr* 1999; 38: 13-126.
11. Aymerich MT, Garriga M, Monfort JM, Nes I, Hugas M. Bacteriocin-producing *Lactobacilli* in Spanish-style fermented sausages: characterisation of bacteriocins. *Food Microbiol* 2000; 17: 33-45.
12. Leenhouts KJ, Kok J, Venema G. Stability of Integrated Plasmids in the Chromosome of *Lactococcus lactis*. *Appl Environ Microbiol* 1990; 56: 2726-2735.
13. de Jong A, van Heel A J, Kok J, et al. BAGEL2: mining for bacteriocins in genomic data. *Nucleic Acids Res* 2010; 38: W647-W651.
14. Maldonado A, Ruiz- Barba JL, Jimenez-Diaz R. Purification and genetic characterization of plantaricin NC8, a novel coculture -inducible two - peptide bacteriocin from *Lactobacillus plantarum* NC8. *Appl Environ Microbiol* 2003; 69: 383-389.
15. Rojo-Bezares B, Sáenz Y, Navarro L, Zarazaga M, Ruiz-Larrea F, Torres C. Coculture-inducible bacteriocin activity of *Lactobacillus plantarum* strain J23 isolated from grape must. *Food Microbiol* 2007; 24: 482-491.

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